

# **STUDIES ON PLANKTON COMMUNITIES OF FISHING HARBOUR ; BAY OF BENGAL, VISA KHAPATNAM, EAST COAST OF INDIA.**

Project submitted to the Andhra University for partial fulfilment for the award

of the degree of

**MASTER OF SCIENCE IN ZOOLOGY**

Presented by

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UNDER THE GUIDANCE OF

**DR.V. RATNA BHARTHI , MSc., M.Phil., Ph.D**



**DEPARTMENT OF ZOOLOGY**

**DR .V.S KRISHNA GOVERNMENT DEGREE & PG COLLEGE(A)**

**VISA KHAPATNAM -- ANDHRA PRADESH**



**Dr.V.S.KRISHNA GOVT. DEGREE COLLEGE**

**(An Autonomous Institution Affiliated to Andhra University)**

**Reaccredited by NAAC with 'A' Grade(3rd Cycle)**

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DEPARTMENT OF ZOOLOGY  
DR V. S. KRISHNA GOVERNMENT DEGREE & PG COLLEGE(A)  
VISAKHAPATNAM



**CERTIFICATE**

This is to certify that the project work entitled **“STUDIES ON PLANKTON COMMUNITIES OF FISHING HARBOUR” : BAY OF BENGAL, VISAKHAPATNAM, EAST COAST OF INDIA** , submitted to the Department Of Zoology, DR V. S. Krishna Government Degree & Pg College (Autonomous) Visakhapatnam is a bona fide record of the research work carried out by **KATARI ANUSHA** of degree of Master of Science in Zoology.

EXAMINAR

P.G COORDINATOR

Dr. T. SAMUEL DAVID RAJ

**DECLARATION**

Hereby declare that the dissertation titled **“STUDIES ON PLANKTON COMMUNITIES OF FISHING HARBOUR; BAY OF BENGAL, VISAKHAPATNAM, EAST COAST OF INDIA”** is submitted to the department of Zoology, DR V S Krishna Government Degree & PG college in partial fulfilment of the requirement for the award of the M.Sc., Degree in Zoology. The work is original and has not been submitted in part or full for any other diploma or degree to this or any other university.

Visakhapatnam

Date:

KATARI ANUSHA

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## ACKNOWLEDGEMENTS

I take this opportunity to sincerely thank the Principal of Dr V.S. Krishna Govt. Degree and PG college, Visakhapatnam , Dr I. Vijaya babu for his immense support to carryout our degree in this college.

I sincerely thank **DR P. JAYA M.Sc., Ph.D.**, Head of the Department of Zoolog , **Dr V S KRISHNA GOVERNMENT DEGREE & PG COLLEGE VISAKHAPATNAM**, for constant encouragement and kind co-operation which enabled me to complete my work successfully.

I express my gratitude to **DR T .SAMUEL DAVIDRAJ , M.Sc, Ph.D., the PG Co ordinator of the department of Zoology** for the constant support rendered throughout the study period.

I would like to express my deep and sincere gratitude to my guide **DR.V.RATNA BHARATHI, Msc.,MPhil.,Ph.D,** Lecturer in Zoology, Department of Zoology for suggesting the project work and guiding throughout the working period which helped me to complete the work in time.

I also extend my thankfulness to fishing harbour employees for giving us opportunity to conduct project.

## **ABSTRACT:**

The present work on Plankton communities of fishing harbour was carried out for a period of 2 weeks at regular intervals revealed three major types of communities namely phytoplankton, proto zooplankton and meta zoo plankton. Of these 18 species are of phytoplankton, 9 species belonging to proto zoo plankton and 4 species belonging meta zoo plankton were observed during the study. The methods involved in collection of samples and qualitative analysis of the samples were discussed detailed manner in the project.

# **1. INTRODUCTION**

The name plankton is derived from the Greek word Plankton which means Wanderer [OR] Drifter . Plankton are the diverse collection of organisms found in water that are unable to propel themselves against a current [or] wind. The individual organisms constituting plankton are called Plankters. In the ocean, they provide a crucial source of food to many small and large aquatic organisms, such as Bivalves, Fish And Whales.

## **MARINE PLANKTON;**

Includes bacteria, archaea, algae, protozoa and drifting [or] floating animals that inhabit the salt water of oceans and brackish water of estuaries. Fresh water plankton are similar to Marine water plankton, but are found in lakes and rivers. Many planktonic species are Microscopic in size , plankton includes organisms over a wide range of sizes, including large organisms such as Jelly fish. This is because plankton are defined by their Ecological Niche and levels of Motility rather than by any Phylogenetic [or] Taxonomic Classification. Technically the term does not include organisms on the surface of the water is called as Neuston [or] those that swim actively in the water called Nekton. Which is composed of strong swimming animals and form benthos. Which includes sessile , creeping.

## **ROLE OF PLANKTON;**

Marine planktons are significant in maintaining the balance & health of the water body and their complex food webs. Nutrients, oxygen and biomass it produces also sustains the terrestrial ecosystem. On the basis of various trophic levels and characteristics, planktons are classified as;

1. PHYTOPLANKTON
2. ZOOPLANKTON

## **1.PHYTO PLANKTON**

Phyto plankton are the Autotrophs [self feeding] components of plankton community and a key part of ocean and fresh water ecosystems. The names comes from the Greek word Phyton Which means Plant and plankton meaning Wander or Drifter. Phytoplankton obtain their energy through the Phyto synthesis as do trees and other plants on land. This means phytoplankton must have light from the sun, so they live in the well-lit surface layers called as Euphotic Zone of oceans and lakes. In comparison with terrestrial plants, phytoplankton are distributed over a large surface area, are exposed less seasonal variation and have markedly faster turnover rates than trees. As a result phytoplankton respond rapidly an a global scale to climatic variations. Phytoplankton form the base of marine and fresh water food webs and are key players in the global Carbon Cycle. They account for about half of global photosynthetic activity and at least of the oxygen production despite amounting to about 1% of the global plant biomass. Most phytoplankton are too small to be individually seen with the Unaided eye. However, when present in high enough numbers. Some varieties may be noticeable as clored patches on the water surface due to presence of Chlorophyl within their cells and accessory pigments such as Phycobiliproteins or Xanthophylls. The term plankton was coined by Victor Hensen in 1887, denotes collectively all free floating and suspended bodies, both plants and animals ,living or dead ,that essentially move passively in a body of water. The phytoplankters are the microscopic plant life of the sea, which constitute the primary producers synthesize the basic food. It belongs to the class algae, which besides chlorophylls posses other characteristics pigments.

The important components of phytoplankton are Diatoms, phytoflagellates [Dinoflagellates], Blue- green algae. Phytoplankton nannochloropsis In addition



to these, two other classes namely Silico flagellates Coccolithophores also belong to the category of phytoplankton. In the sea water, the diatoms and dinoflagellates are the more obvious representative of the phytoplankton in terms of both cell size and availability when water sample are examined under microscope. Depending on size on the size grading of phytoplankton organisms they are grouped under:

1. More than 1mm : Macroplankton
2. less than 1mm : Microplankton
3. Organisms of 5-50um : Nano plankton
4. Less than 5 um : Ultra plankton
5. less than 1 um : Pico plankton

### **Characteristics of phytoplankton:**

Most phytoplankton organisms are unicellular. The larger colonial forms possess individual cells that are usually of uniform structure and appearance. Some planktonic green algae are of filamentous organisms organisation, and in some and dinoflagellates chains of loosely certain cell characteristics and ease of identification will clearly be limited by cell size and the degree of magnification obtainable. For identification purpose we look for the following with phytoplankton organisms: 1. Cell shape 2. Cell dimension 3. Cell wall 4. Mucilage layer 5. Chloroplast 6. Flagella and 7. Reserve substance.

## **Types of phytoplankton;**

Based on cell wall arrangement and cell structure the phytoplankton are classified into five types ;

1. Diatoms
2. Dinoflagellates
3. Blue green algae
4. filamentous

## **DIATOMS**

A diatom is any member of a large group comprising several genera of algae, specifically micro algae, found in oceans, water ways and soils of the world. Living diatoms make up a significant portion of earth's Biomass. Diatoms are Unicellular organisms they occur either as solitary cells or in Colonies. Which can take the shape of ribbons, fans, stars.

## **Morphological characters of diatoms**

Diatoms are single -celled organisms which secrete Intricate Skeleton .These may be elongate with a bilateral plane of symmetry [or] they may be round and radially symmetrical. The skeleton of a diatom [or] frustule is made of very pure silica coated with a layer of organic material. The skeleton is divided into Epitheca& Hypotheca. Pennate diatoms show a long slit, the Raphe, along the axis. Within their silica walls'Diatoms shows a typical level of Eukaryotic Organization.

## **DINOFLAGELLATES**

The dinoflagellates are a Monophyletic group of single celled eukaryotes constituting the phylum Dinoflagellate. Many dinoflagellates are photosynthetic but a large fraction of these are in fact Mixotropic,

combining photosynthesis with ingestion of prey. Some species are Endosymbionts of marine animals and play an important part in the biology of Coral reefs.

## **MORPHOLOGICAL CHARACTERS OF DINOFLAGELLATES**

Dinoflagellates are unicellular and possess two dissimilar flagella arising from the ventral side. They have a ribbon like transverse flagellum with multiple waves that beats to the cell's left and a more conventional one. Dinoflagellates have a complex cell covering called an Amphiesma [OR] Cortex compared of series of membranes flattened vesicles called Alveoli.

Examples;-

Noticula, Gonyalax

## **BLUE GREEN ALGAE**

Cyanobacteria also known as cyanophyta are a phylum of Gram Negative Bacteria that obtain energy via Photosynthesis. Cyanobacteria use Photosynthetic pigments such as Carotenoids, phycobilin and various forms of chlorophyll which absorb energy from light. These have flattened sacs called Thylakoids where photosynthesis is performed. Cyanobacteria are the first organisms known to have produced oxygen.

## **MORPHOLOGICAL CHARACTERS OF BLUE GREEN ALGAE**

These are unicellular and filamentous to colonial forms. Many cyanobacteria form motile filaments of cells called Hormogonia that travel away from the main biomass to bud and form new colonies elsewhere. To break away from the parent colony, a hormogonium often must part a weak cell in a filament called Necridium.

Examples;- Anabaena ,Nostoc

## **FILAMENTOUS ALGAE**

The name algae refers to broad and immensely diversely group of photosynthetic eukaryotic life forms. These organisms are not related since they do not have a common ancestor. Algae can be found in a variety of habitat , including seas, rivers, and lakes as well as ponds, brackish water and even snow. Algae are often green in colour however they can come in a range of different colours. Filamentous algae are microscopic algae that create threads or mesh like filaments by linking themselves.

### **EXAMPLES ;**

Spirogyra and Ulothrix are examples of filamentous algae. Spirogyra is free floating green algae that can be found in ponds,lakes, and other fresh water environments. The vegetative structure of these algae is filamentous and unbranched. Ulothrix is a filamentous green algae genus found in both fresh and salt water.

## **ZOO PLANKTON**

Zooplankton are the animal component of the planktonic community . ZOO comes from the Greek word for animal. Zooplankton can be contrasted with phytoplankton which are the plant component of the plankton community. Zooplankton are Heterotrophic. Whereas phytoplankton are Autotrophic [Self Feeding]. In other wordzooplankton connect manufacture their own food. Rather, they must eat other plants or animals instead. Most zooplankton are Microscopic but some [such as jelly fish] are Macroscopic. Zooplankton can be classified into 2 types

1. Micro Zooplankton
2. Macro Zooplankton

## **MICRO ZOO PLANKTON**

Micro zooplankton are defined as Heterotrophic and Mixotrophic plankton. They are primarily consists of Phagotrophic Protists including Ciliates, Dinoflagellates. As the primary consumers of marine phytoplankton, micro zooplankton consume 59-75% daily of the marine primary production. Despite their ecological importance micro zooplankton remain understudied. Routine oceanographic observations seldom monitor microzooplankton biomass or herbivory rate, although the dilution technique, an elegant method of measuring micro zooplankton herbivory rate has been developed for almost four decades [Landry and Hassett 1982]. The number of observation of microzooplankton herbivory rate is around 1600 globally far less than that of primary productivity [ $>50,000$ ]. Micro zooplankton are also pivotal regenerators of nutrient which fuel primary production and food sources of metazoans. Micro zooplankton can be classified into 3 major types. 1. Zoo flagellates 2. Ciliates 3. Copepods of which ciliates and copepods are predominant forms of zooplankton.

### **1. CILIATES**

Ciliates are a group of protozoans characterized by the presence of hair like structures called cilia. Which are identical in structure to eukaryotic flagellum but are in general shorter and present in much larger numbers with a different undulating patterns. . Cilia occurs in all members of the group and are variously used in swimming, attachment feeding and sensation. Ciliates reproduce asexually by various kinds of fission. Most ciliates are heterotrophs feeding on smaller organisms such as bacteria and algae and detritus swept into the oral groove by modified oral cilia.

## **2. COPEPODS**

Copepods are group of small crustaceans found in nearly every freshwater and saltwater habitat. Copepods are sometimes used as Biodiversity Indicators. Copepods vary considerably but are typically 1 to 2 mm [ $\frac{1}{32}$  to  $\frac{3}{32}$  in] long, with a teardrop shaped body and large intestine. Like other crustaceans but they are so small that in most species, this thin armour and the entire body is almost totally transparent. Most copepods have a single median compound eye, usually bright red and in the centre of the transparent head. Because of their small size, copepods have no need any heart [or] circulatory system and most also lack gills. Instead they absorb oxygen directly into their bodies, their excretory system consists of maxillary glands.

Eg; calanoida, cyclops, cyclopoida

## REVIEW OF LITERATURE

The enormous impact that marine phytoplankton have on global biogeochemical cycles and marine ecosystem functioning largely motivates which aspects of their ecology, physiology and evolution are routinely measured. Often, their diversity is divided into functional groups based on metabolic capacity and impact on biogeochemical cycles (DeLong 2009; Fuhrman 2009).

Phytoplankton form the base of marine food webs and comprised a taxonomically broad range of organisms that includes both prokaryotes and eukaryotes and consists of thousands of species (Falkowski and Raven 2007). The relative contribution of different taxa to primary production influences the export of organic carbon to the deep ocean, termed the ‘biological pump’ (Volk and Hoffert 1985; De La Rocha and Passow 2007). Different taxa differ in size and composition of their cell walls and coverings, which influence their sinking rates and thus their impact on the biological pump. For example, the smallest constituents of the marine phytoplankton are the picoplankton (0.2–2  $\mu\text{m}$ ), which include taxa such as cyanobacteria and prasinophytes, and sink slowly, at rates of  $< 0.5$  m/day (Bach et al. 2012). In contrast, larger cells such as diatoms are covered in biogenic silica, which acts as ballast, and accelerates their sinking to rates of up to 35 m/day (Miklasz and Denny 2010). Coccolithophores are commonly covered with calcium carbonate plates called ‘coccoliths’, which also acts as a ballasting agent, yielding sinking rates of nearly 5 m/day (Bach et al. 2012). In addition to ballast material, processes such as aggregation and the packaging of small cells into large faecal pellets by grazers also affects sinking rates and the efficiency of the biological pump (Klaas and Archer 2002; De La Rocha and Passow 2007).

Given their phylogenetic breadth, different phytoplankton taxa will likely have different responses to climate change. For example, it has been suggested that

decreased ocean pH may hinder the ability of coccolithophores to generate calcareous plates, potentially influencing their impact on the biological pump (Riebesell et al. 2000; Langer et al. 2009; Hoppe et al. 2011). Though tiny, coccolithophores are numerous enough for this to matter, with blooms of the cosmopolitan species *Emiliana huxleyi* exceeding 250,000 km<sup>2</sup>. When these cells die and release their coccoliths, the remains of these blooms can be seen from space (Holligan et al. 1993; Balch et al. 1996; Tyrrell and Merico 2004). Because reduced calcification is thought to affect fitness in calcifying coccolithophores (Beaufort et al. 2011), there has been particular interest in the possibility that adaptive evolution may mitigate the reduction in calcification as oceans acidify. There is evidence that this can sometimes occur, since well-calcified morphotypes of coccolithophores have been found in low pH regions of the ocean, despite a general trend of decreasing calcification with rising pCO<sub>2</sub> levels, though part of this indicates species succession rather than evolution within species (Beaufort et al. 2011).



## **STUDY AREA AND METHODOLOGY**

### **STUDY AREA;**

The present study was carried out in the Fishing Harbour of Visakhapatnam for the collection of plankton organisms. Fishing harbour located on the city's east coast is one of the India's largest fishing harbour offering a spectacular view of Bay of Bengal. The fishing harbour spread over an area of 26 hectares and it was established in the year 1976 and is managed by the Visakhapatnam port trust. Its capacity is 700 mechanised boats and 300 beach landing crafts. Visakhapatnam port consists of three harbours - outer harbour, inner harbour and the fishing harbour. The outer harbour has 6 berths and it capable for handling vessels with a draft upto 17 meters and the smaller inner harbour has 18 berths those are paramax compatible. The dolphins nose hill to the north of the entrance channel protects the harbour from cyclones that can strike the east coast.

Fishing harbour has easy access for the public as it has big market area for commercial purpose of exports and other marketing needs. There are many fishing crafts placed in jetties for active fishing purposes. During the study Jetty 6 was taken for sample collection as this jetty is free from the market area of the harbour and the fishing vessel matsya shikari of FSI berths at this jetty. The crew of the vessel are supportive during the sample collection and the water is undisturbed in this region.

## Location Map for Fishing Harbour Vizag



PLATE 1: Fig a) The google map showing fishing harbour area

Fog)

## METHODOLOGY



During the study the surface water samples are collected from the fishing harbour area entrance channel of Visakhapatnam harbour. A plastic bucket was tied with roap and bucket full of water is collected during morning hours of day for a period of 10 days (every 24 hours same ). The samples were brought to the

laboratory for analysis and identification. 1 litre of sample is poured in 1000 ml measuring cylinder for quantitative observation of plankton. Another 1 litre is filtered through whatman filter paper GF/C using suction apparatus. Top 100ml of filtrate is collected in beaker for quantitative observation of species composition and identification. The filtered water sample is observed under the microscope by taking a drop of the sample on a glass slide while observing the species we used 10x20x40 magnified lens (Labomed 5100 model). The samples are stained with vital stains such as methyl green, neutral red and lugol's solution. For quantitative analysis 1000ml of water was taken in cylinder measuring jar to which 10 ml of lugol's iodide is added. This helped in fixing of plankton. After 48 hours supernatant water sample is pipetted out and left over 100ml is preserved in sample bottle. The preserved sample is to observing the species composition in a counting chamber sedgewick rafter counter for enumeration. By following above methods the quantitative & qualitative analysis samples are carried out.

## **RESULTS**

The present study undertaken in fishing harbour area for a period of 2 weeks to understand species composition and distribution of plankton communities. The observation revealed 3 major groups of planktons.

1. Phytoplankton

2. Protozooplankton

3. Metazooplankton

Total of 18 species of phytoplankton, 13 species of protozooplankton and 4 species of metazooplankton identified from the sample.

**Table 1: Plankton community composition in fishing harbour during short term study:**

<u>S.no</u>	<b>Plankton communities</b>	<u>Number of species</u>
1.	Phytoplankton	18
2.	Flagellates	4
3.	Ciliates	9
4.	Copepods	4

**Table 2: Phytoplankton occurrence at 24 hr intervals during the study at fishing harbour.**

S no	Name	Day 1	Day 2	Day 3	Day 4	Day 5	Day 6	Day 7
1.	<i>Navicula sp</i>	+	+	+	+	+	+	+
2.	<i>Skeletonema sp</i>	+	+	-	+	-	+	+
3.	<i>Pleurosigma sp</i>	+	+	+	+	+	+	+

4	<i>Chaetoceros sp.</i>	+	+	+	+	+	+	+
5	<i>Nitschizia longissima sp</i>	-	-	+	+	-	+	+
6	<i>Thalassiosera pseudonana</i>	-	-	-	-	+	+	+
7	<i>Dytilium sp</i>	-	-	-	-	-	+	+
8	<i>Cymbella sp</i>	-	-	-	-	-	+	+
9	<i>Blue green algae</i>	-	-	+	+	-	+	+
10	<i>Oscillatoria sp</i>	-	-	-	+	+	+	+

## 1. *Navicula* sp.

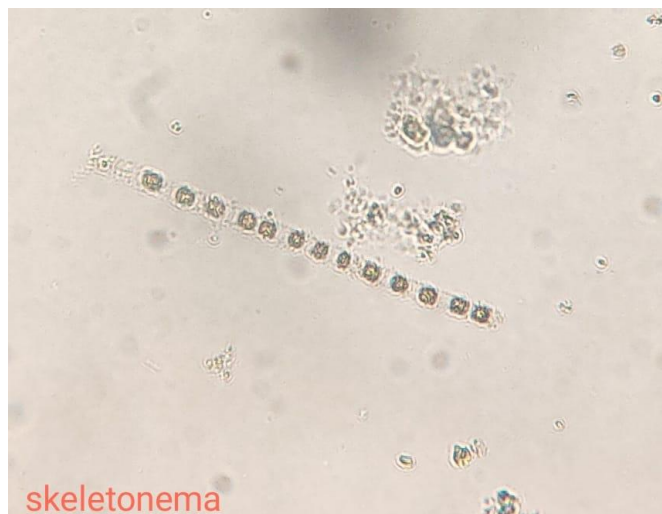


Domain:	<a href="#">Eukaryota</a>
Clade:	<a href="#">Diaphoretickes</a>
Clade:	<a href="#">SAR</a>
Clade:	<a href="#">Stramenopiles</a>
Phylum:	<a href="#">Gyrsta</a>
Subphylum:	<a href="#">Ochrophytina</a>
Class:	<a href="#">Bacillariophyceae</a>
Order:	<a href="#">Naviculales</a>
Family:	<a href="#">Naviculaceae</a>
Genus:	<i>Navicula</i> <a href="#">Bory de Saint-Vincent</a> , 1822

## DESCRIPTION OF GENERA;

*Navicula* is a genus of boat-shaped diatom algae, comprising over 1,200 species. *Navicula* is Latin for small ship and also a term in English for a boat-shaped incense-holder. Diatoms are eukaryotic, primarily aquatic, single-celled photosynthetic organisms play an important role in global ecology, producing about a quarter of all the oxygen within Earth's biosphere, often serving as foundational organisms, or keystone species in the food chain of many environments where they provide a staple for the diets of many aquatic species. *Navicula* diatoms have been observed to possess a motile ability to glide over one another and on hard surfaces such as microscope slides. Around the outside of the navicula's shell is a girdle of mucilage strands that can flow and thus act as a tank track.

### 2. *Skeletonema* sp



Domain:	<a href="#">Eukaryota</a>
Clade:	<a href="#">Diaphoretickes</a>
Clade:	<a href="#">SAR</a>
Clade:	<a href="#">Stramenopiles</a>

Phylum:	<a href="#">Gyrsta</a>
Subphylum:	<a href="#">Ochrophytina</a>
Class:	<a href="#">Bacillariophyceae</a>
Order:	<a href="#">Thalassiosirales</a>
Family:	<a href="#">Skeletonemataceae</a>
Genus:	<i>Skeletonema</i> <a href="#">R. K. Greville</a> , 1865

## DESCRIPTION OF GENERA;

*Skeletonema* is a genus of diatoms in the family Skeletonemataceae. It is the type genus of its family. The genus *Skeletonema* was established by R. K. Greville in 1865 for a single species, *S. barbadense*, found in the Barbados deposit [Jung 2009]. These diatoms are photosynthetic organisms, meaning they obtain carbon dioxide from their surrounding environment and produce oxygen along with other by products. Reproduce sexually (sexual reproduction is oogamous) and asexually [Guiry 2011]. *Skeletonema* sp. belong to the morphological category referred to as centric diatoms. These are classified by having valves with radial symmetry and the cells lack significant motility [Horner 2002]. *Skeletonema* are cylindrical shaped with a silica frustule. Cells are joined by long marginal processes to form a filament [Horner 2002]. Their length ranges from 2-61 micrometers, with a diameter ranging from 2-21 micrometers [Hasle 1997].



### 3. PLEUROSIGMA



Domain:	<a href="#">Eukaryota</a>
Clade:	<a href="#">Diaphoretickes</a>
Clade:	<a href="#">SAR</a>
Clade:	<a href="#">Stramenopiles</a>
Phylum:	<a href="#">Gyrista</a>
Subphylum:	<a href="#">Ochrophytina</a>
Class:	<a href="#">Bacillariophyceae</a>
Order:	<a href="#">Naviculales</a>
Family:	<a href="#">Pleurosigmataceae</a>
Genus:	<i>Gyrosigma</i> <a href="#">A.H.Hassall</a> , 1845

## DESCRIPTION OF GENERA;

The genus of marine planktonic organisms known as *Pleurosigma* is named so for its shape. *Sigma* is a latin root used in reference to the shape of the organism. It has also been related to the orbital movement of electrons around an internuclear axis, which is similar to the orbital movement the Pleurosigma diatom makes around one of its tips during movement. The genus employs a pulsating of the body to induce gliding movement as well as occasional circular rotations around one end of the organism to change course.

### 4. *Chaetoceros* sp.



Domain:	<a href="#">Eukaryota</a>
Clade:	<a href="#">Diaphoretickes</a>
Clade:	<a href="#">SAR</a>
Clade:	<a href="#">Stramenopiles</a>
Phylum:	<a href="#">Gyrista</a>
Subphylum:	<a href="#">Ochrophytina</a>
Class:	<a href="#">Bacillariophyceae</a>
Order:	<a href="#">incertae sedis</a>
Family:	<a href="#">Chaetocerotaceae</a>
Genus:	<i>Chaetoceros</i> Ehrenberg (1844)

## DESCRIPTION OF GENERA;

*Chaetoceros* consists of cells linked together, forming long chains. Individual cells are elliptical to circular in valve view, making them centric diatoms, and are rectangular in girdle view. Like other diatoms, cells of *Chaetoceros* are surrounded by siliceous cell walls known as frustules. Each frustule has four hollow processes called setae, or spines, that allow adjacent cells to link together and form colonies. Colonies can form chains that are coiled, straight, or curved. Cell size can range from <10 um to 50 um. Some species of *Chaetoceros* produce resting spores that are highly tolerant to adverse conditions.

## 5. *Nitzschia longissima*



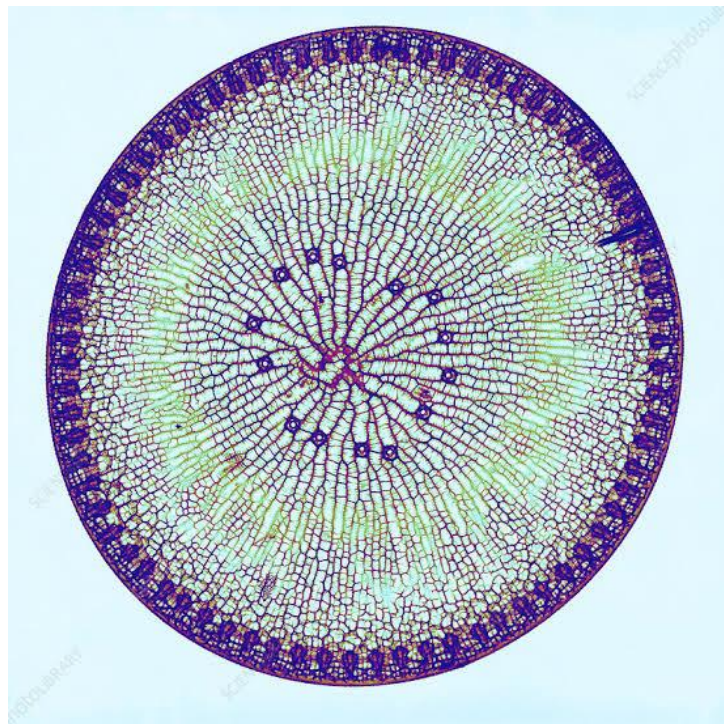
Domain:	<a href="#">Eukaryota</a>
Clade:	<a href="#">Diaphoretickes</a>
Clade:	<a href="#">SAR</a>
Clade:	<a href="#">Stramenopiles</a>
Phylum:	<a href="#">Gyrista</a>
Subphylum:	<a href="#">Ochrophytina</a>
Class:	<a href="#">Bacillariophyceae</a>
Order:	<a href="#">Bacillariales</a>
Family:	<a href="#">Bacillariaceae</a>

## DESCRIPTION OF GENERA;

*Nitzschia* sp. is a common pennate marine diatom. In the scientific literature, this genus, named after Christian Ludwig Nitzsch, is sometimes referred to incorrectly as **Nitzchia**, and it has many species described, which all have a similar morphology.

In its current circumscription, *Nitzschia* is paraphyletic. *Nitzschia* is found mostly in colder waters, and is associated with both Arctic and Antarctic polar sea ice, where it is often found to be the dominant diatom. *Nitzschia* includes several species of diatoms known to produce the neurotoxin known as domoic acid, a toxin responsible for the human illness called amnesic shellfish poisoning. The species N. frigida is found to grow exponentially even at temperatures between  $-4$  and  $-6$  °C.

## 6. *Thalassiosira pseudonana*



Superphylum:	<a href="#">Heterokonta</a>
Class:	<a href="#">Coscinodiscophyceae</a>
Order:	<a href="#">Thalassiosirales</a>
Family:	<a href="#">Thalassiosiraceae</a>
Genus:	<a href="#">Thalassiosira</a>
Species:	<b><i>T. pseudonana</i></b>

## DESCRIPTION OF GENERA;

*Thalassiosira pseudonana* is a species of marine centric [diatoms](#). It was chosen as the first [eukaryotic](#) marine [phytoplankton](#) for whole [genome sequencing](#). *T. pseudonana* was selected for this study because it is a model for diatom physiology studies, belongs to a genus widely distributed throughout the world's oceans, and has a relatively small genome at 34 mega [base pairs](#). Scientists are researching on diatom light absorption, using the marine diatom of *Thalassiosira*. The diatom requires a high enough concentration of CO<sub>2</sub> in order to utilize C<sub>4</sub> metabolism (Clement *et al.* 2015).

## 7. *Dityllum brightwellii*

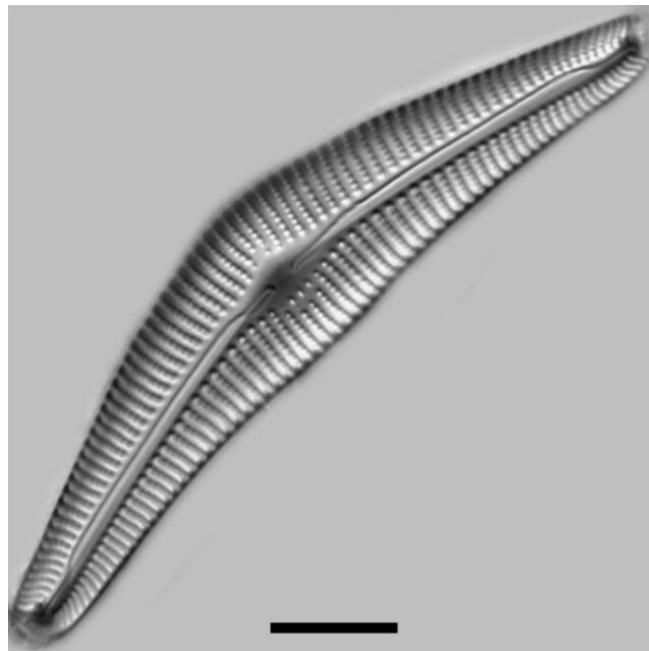


Domain:	<a href="#">Eukaryota</a>
Clade:	<a href="#">Diaphoretickes</a>
Clade:	<a href="#">SAR</a>
Clade:	<a href="#">Stramenopiles</a>
Phylum:	<a href="#">Gyrista</a>
Subphylum:	<a href="#">Ochrophytina</a>
Class:	<a href="#">Bacillariophyceae</a>
Order:	<a href="#">Lithodesmiales</a>
Family:	<a href="#">Lithodesmiaceae</a>
Genus:	<a href="#">Ditylum</a>
Species:	<b><i>D. brightwellii</i></b>

## DESCRIPTION OF GENERA;

*Ditylum brightwellii* is a species of cosmopolitan marine centric [diatoms](#). It is a [unicellular photosynthetic autotroph](#) that has the ability to divide rapidly and contribute to spring phytoplankton blooms. The *D. brightwellii* [cell](#) has a high length to diameter ratio. The cell wall is silicified, as is characteristic of all diatoms. This hard, porous covering is known as the [frustule](#) and causes the cell to be more dense than the surrounding water. Oceanic currents and surface winds prevent *D. brightwellii* cells from sinking beneath the [euphotic zone](#). Cells range in size from 25-100µm in diameter and 80-130µm in length. The valve is most often triangular in shape, but can also be biangular or quadrangular. A long hollow tube called the rimoportula is located centrally and extends from each valve.

## 8. CYMBELLA





Domain:	<a href="#">Eukaryota</a>
Clade:	<a href="#">Diaphoretickes</a>
Clade:	<a href="#">SAR</a>
Clade:	<a href="#">Stramenopiles</a>
Phylum:	<a href="#">Gyrista</a>
Subphylum:	<a href="#">Ochrophytina</a>
Class:	<a href="#">Bacillariophyceae</a>
Order:	<a href="#">Cymbellales</a>
Family:	<a href="#">Cymbellaceae</a>
Genus:	<i>Cymbella</i> <a href="#">C.Agardh</a> , 183

## DESCRIPTION OF GENERA;

Cymbella is a diatom genus in the family cymbellaceae including over 800 species. Valves slightly to strongly asymmetric to the apical axis. Distal raphe ends deflected dorsally. Apical porefields present. Stigmata if present located on the ventral side. Chloroplast usually a single double H with a central pyrenoid, pyrenoid lying on the dorsal side of the girdle. Cymbella lanceolata has a single chloroplast with a central pyrenoid and many lobes coming off in different directions.

## 9. BLUE GREEN ALGAE FILAMENTS



Domain:	<a href="#">Bacteria</a>
Clade:	<a href="#">Terrabacteria</a>
(unranked):	<a href="#">Cyanobacteria-Melainabacteria group</a>
Phylum:	Cyanobacteria <a href="#">Stanier</a> , 1973
Class:	Cyanophyceae

## DESCRIPTION OF GENERA;

**Cyanobacteria** also called **Cyanophyta**, are a [phylum](#) of [gram-negative bacteria](#)[\[4\]](#) that obtain energy via [photosynthesis](#). The name *cyanobacteria* refers to their color . which similarly forms the basis of cyanobacteria's common name, **blue-green algae**, although they are not usually scientifically classified as [algae](#). They appear to have originated in a freshwater or terrestrial environment. Sericytochromatia, the proposed name of the [paraphyletic](#) and most basal group, is the ancestor of both the non-photosynthetic group [Melainabacteria](#) and the photosynthetic cyanobacteria, also called Oxyphotobacteria. Cyanobacteria use [photosynthetic pigments](#), such as [carotenoids](#), [phycobilins](#), and various forms of [chlorophyll](#), which absorb energy from light. Unlike [heterotrophic](#) prokaryotes, cyanobacteria have [internal membranes](#). These are flattened sacs called [thylakoids](#) where photosynthesis is performed.

## 10.OSCILLATORIA SPECIES



Domain: [Bacteria](#)

Phylum: [Cyanobacteria](#)

Class: [Cyanophyceae](#)

Order: [Oscillatoriales](#)

Family: [Oscillatoriaceae](#)

Genus: *Oscillatoria*  
[Vaucher](#) ex [Gomont](#),  
1822

## DESCRIPTION OF GENERA;

*Oscillatoria* is a genus of [filamentous cyanobacterium](#) which is often found in freshwater environments, such as hot springs, and appears blue-green.<sup>[1]</sup> Its name refers to the [oscillating](#) motion of its filaments as they slide against each other to position the colony facing a light source.<sup>[2]</sup> *Oscillatoria* reproduces by [fragmentation](#), facilitated by dead cells which separate a filament into separate sections, or [hormogonia](#), which then grow.<sup>[1]</sup> *Oscillatoria* uses photosynthesis to survive and reproduce. Each filament of *oscillatoria* consists of trichome <sup>[clarification needed]</sup> which is made up of rows of cells. The tip of the trichome oscillates like a pendulum. In reproduction, it takes place by vegetative means only. Usually the filament breaks into a number of fragments called hormogonia. Each hormogonium consist of one or more cells and grow into a filament by cell division in one direction.

# CILIATES

## 1. STROMBIDIUM

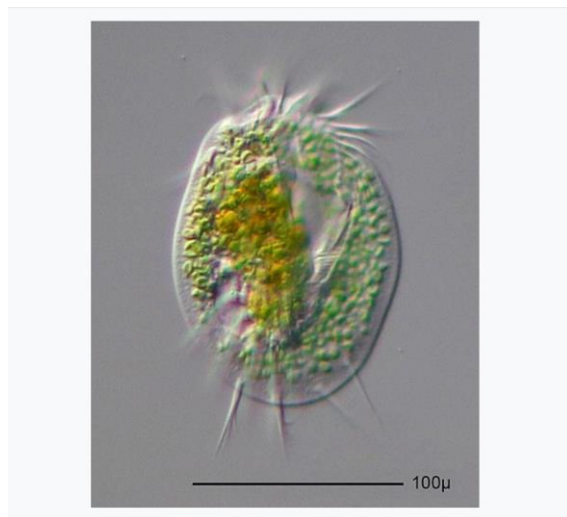


Domain:	<a href="#">Eukaryota</a>
(unranked):	<a href="#">Sar</a>
(unranked):	<a href="#">Alveolata</a>
Phylum:	<a href="#">Ciliophora</a>
Class:	<a href="#">Spirotrichea</a>
Subclass:	<a href="#">Oligotrichia</a>
Order:	<a href="#">Oligotrichida</a>
Family:	<a href="#">Strombidiidae</a>
Genus:	<a href="#">Strombidium</a>
Species:	<b><i>S. lagenula</i></b>

## DESCRIPTION OF GENERA;

*Strombidiumlagenula* is a species of [marine planktonic ciliates](#) belonging to the genus *Strombidium* (which contains about 31 species) in the order [Oligotrichida](#). Like other ciliates, they are unicellular eukaryotes ([protists](#)) that move using [cilia](#). However in *Strombidiumlagenula* and oligotrichs in general, the cilia on the cell body have been either lost or heavily reduced to "bristles", while the cilia surrounding the oral opening form large and prominent structures called [membranelles](#), which are also used for locomotion.

## 2.EUPLOTES



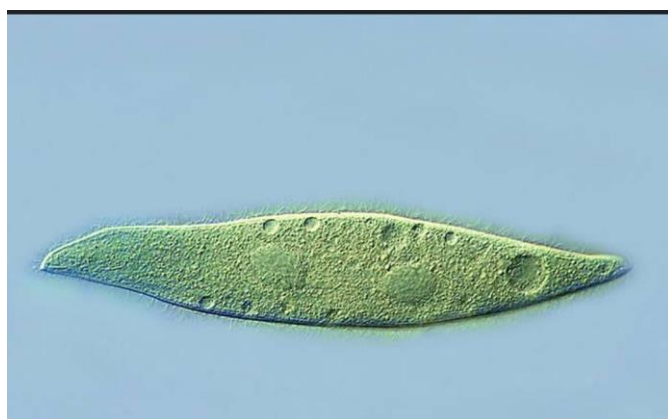
Domain:	<a href="#">Eukaryota</a>
(unranked):	<a href="#">SAR</a>
(unranked):	<a href="#">Alveolata</a>
Phylum:	<a href="#">Ciliophora</a>
Class:	<a href="#">Spirotrichea</a>
Subclass:	<a href="#">Euplotia</a>
Order:	<a href="#">Euplotida</a>

Suborder:	<a href="#">Euplotina</a>
Family:	<a href="#">Euplotidae</a>
Genus:	<i>Euplotes</i>
	<a href="#">Ehrenberg</a> , 1830

### DESCRIPTION OF GENERA;

*Euplotes* cells are inflexible, [dorsoventrally](#) flattened, and roughly ovoid, with a very large oral region (peristome) bordered on the left by a long "adoral zone of [membranelles](#)" (AZM). Like other [spirotrich](#) ciliates, *Euplotes* move and feed with the help of compound ciliary [organelles](#) called "cirri," made up of thick tufts of [cilia](#) sparsely distributed on the cell. Strong cirri on the [ventral](#) surface of the cell enable *Euplotes* to walk or crawl on submerged detritus and vegetation. All species of *Euplotes* have a group of stiff bristles ([caudal](#) cirri), which protrude from the posterior of the cell.

### 3.AMPHILECTUS



Kingdom: [Animalia](#)

Phylum: [Porifera](#)

Class: [Demospongiae](#)

Order: [Poecilosclerida](#)

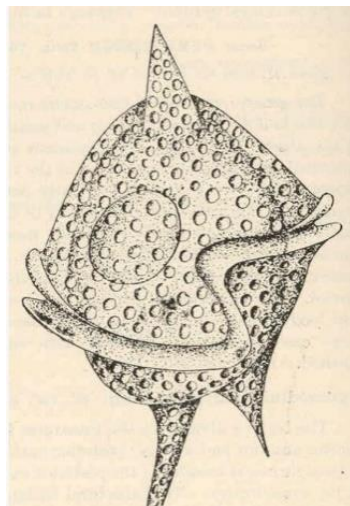
Family: [Esperiopsidae](#)

Genus: *Amphilectus*  
Vosmaer,  
1880<sup>[1]</sup>

### DESCRIPTION OF GENERA;

Body laterally compressed, highly elongate with anterior neck-like region which bends towards the dorsal edge. Oral aperture a slit on convex edge of neck region, extends less than halfway down the body. Ciliation present on both lateral surface although there is a tendency to some reduction on the left surface resulting in it being difficult to distinguish. Ciliation on right surface is extensive and forms longitudinal rows which converge on each other in the anterior region. There is a distinctive area of cilia along the oral slit forming a mane-like brush. Trichocysts commonly present particularly in neck.

### 4.PERIDINIUM





Domain:	<a href="#">Eukaryota</a>
Clade:	<a href="#">Diaphoretickes</a>
Clade:	<a href="#">SAR</a>
Clade:	<a href="#">Alveolata</a>
Phylum:	<a href="#">Myzozoa</a>
Superclass:	<a href="#">Dinoflagellata</a>
Class:	<a href="#">Dinophyceae</a>
Order:	<a href="#">Peridiniales</a>
Family:	<a href="#">Peridiniaceae</a>
Genus:	<i>Peridinium</i> <a href="#">Ehrenberg</a>

## DESCRIPTION OF GENERA;

*Peridinium* is a genus of motile, [marine](#) and [freshwater dinoflagellates](#). Their morphology is considered typical of the armoured dinoflagellates, and their form is commonly used in diagrams of a dinoflagellate's structure. *Peridinium* can range from 30 to 70 µm in diameter, and has very thick [thecal](#) plates.

## CONCLUSION

The short term study on Plankton communities revealed interesting observations in the fishing harbour of Visakhapatnam. Plankton communities are very important in establishing the productivity of ecosystem. The marine phytoplankton act as producers of marine ecosystem. The fauna of marine ecosystem depend on phytoplankton productivity During the study results showed ;

18 phytoplanktons,9 protozooplankton,4 copepodes respectively. The phytoplankton are the primary producers and zooplankton act as primary consumers of marine ecosystem.

## REFERENCES;

- 1.Ash, C., Stone, R. (2003). A question of dose. Science, 300, 925.
2. Barón, M., Arellano, J.B., Lo pez Gorge, J. (1995). Cu and photosystem II, a controversial relationship. *Physiologia Plantarum*. 94, 174-180.
3. Beleneva, L. A., Kukhlevsky, A. D., Kharchenko, U. V. and Kovalchuk, Y. L. (2011). Resistance to Copper Ions and Antibiotics in Marine Heterotrophic Bacteria in the Coastal Waters of Vietnam. *Russian Journal of Marine Biology*, 37(4), 284-290.
4. Bertrand, M., Schoefs, B., Siffel, P., Rohacek, K. and Molnar, L. (2001). Cadmium inhibits epoxidation of diatoxanthin to diadinoxanthin in the xanthophyll cycle of the marine diatom *Phaeodactylum tricornutum*. *FEBS letters*, 508(1),153-156.

5. Biswas, H. and Bandyopadhyay, D. (2013). Effects of iron availability on pigment signature and biogenic silica production in the coastal diatom *Chaetoceros gracilis*, *Advances in Oceanography and Limnology*, 4(1), 20-4
6. Brand, L E, Sunda, W. G., and Guillard, R. R. L. (1986). Reduction of marine phytoplankton reproduction rates by copper and cadmium. *Journal of Experimental Marine Biology and Ecol*, 96, 225-250.
7. Bruland, K. W., Donat, J. R. and Hutchins, D. A. (1991). Interactive influences of bioactive trace metals on biological production in oceanic waters. *Limnology and Oceanography*, 36, 1555-1577.
8. Caspi, V., Droppa, M., Horváth, G., Malkin, S., Marder, J. B. and Raskin, V. L. (1999). The effect of copper on chlorophyll organization during greening of barley Leaves. *Photosynthesis Research*, 62 (2), 165-174.
9. Chadd, H. E., Newman, J., Mann, N. H., Carr, N. G. (1996). Identification of iron superoxide dismutase and a copper/zinc superoxide dismutase enzyme activity within the marine cyanobacterium *Synechococcus* sp. WH 7803. *FEMS Microbiology Letters*, 138(2-3), 161-5
10. Cid, A., Herrero, C., Torres, E., Abalde, J. (1995). Copper toxicity on the marine microalga *Phacodactylum tricorutum* effects on photosynthesis and related parameters. *Aquatic Toxicology*, 31, 165-174.
11. Coale, K.H., (1991). Effects of iron, manganese, copper, and zinc enrichments on productivity and biomass in the subarctic Pacific. *Limnology and Oceanography*, 36(8), 1851-1864.
12. Croot, P. L., Moffet, J. W. and Brand L. E. (2000). Production of extracellular Cu complexing ligands by eukaryotic phytoplankton in response to Cu stress. *Limnology and Oceanography*, 45(3), 619-627.

13. Davis, A. K., Hildebrand, M., and Palenik, B. (2006). Gene Expression Induced by Copper Stress in the Diatom *Thalassiosira pseudonana*. *Eukaryotic Cell*, 5(7), 1157-1168.
14. Davis, S. A. G. (1966). Studies of the accumulation of radio-iron by a marine diatom, p. 938-991. Zn Radioecological concentration processes. Proc. Int. Symp. Stockholm. Pergamon.
15. De Vos, C. H. R., Schat, H., Vooijs, R. and Ernst, W. A. O. (1989). Copper induced damage to the permeability barrier in roots of *Silene cucubalus*. *Journal of Plant Physiology*, 135, 164-169.
16. Debelius, B., Forja, J.M., DelValls, Á. and Lubián, LM. (2010). Toxic effect of copper on marine picophytoplankton populations isolated from different geographic locations. *Scientia Marina*, 74(S1), 133-141.
17. Florence, T. M. and Stauber, J. L. (1986). Toxicity of copper complexes to the marinediatom *Nitzschia closterium*. *Aquatic Toxicology*, 8, 11-26.
18. Gadd, G. M. and Griffiths, A. J. (1978). Microorganisms and Heavy metal toxicity. *Microbial Ecology*, 4, 303-317.
19. Goulder, R., Blanchard, A. S., Metcalf P. J., and Wright, B. (1979). Inhibition of Estuarine Bacteria by Metal Refinery Effluent. *Marine Pollution Bulletin*, 10, 170-17.
20. Green, L. F., McCarthy, J. M. and King, C. G. (1939), Inhibition of respiration and photosynthesis in *Chlorella pyrenoidosa* by organic compounds that inhibit copper catalysis. *The Journal of Biological Chemistry*, 128, 447-453.
21. Gustavson, K., Petersen, S., Pedersen, B., Stuer-Lauridsen, F., Pedersen, S., Wangberg, S.A.(1999) Pollution-induced community tolerance (PICT) in coastal phytoplankton communities exposure to Cu. *Hydrobiologia*, 416,125-138.

22. Harrison, W. G. Eppley, R. W. and Renge, E. H. (1977). Phytoplankton nitrogen metabolism, nitrogen budgets and observations on copper toxicity, controlled ecosystem pollution experiment. *Bulletin of Marine Science*. 27(1), 44-57.
23. Hasle, G. R., Syversten, E. E. (1997). Marine diatoms. In, Tomas CR (Ed) *Identifyingmarine phytoplankton* (pp. 5-385). Academic Press, San Diego.
24. Henley, S. F., Annett, A. L., Ganeshram, R. S., Carson, D. S., Weston, K., Crosta, X., Tait, A., Dougans, J., Fallick A. E., and Clarke. A. (2012). Factors influencing the stable carbon isotopic composition of suspended and sinking organic matter in the coastal Antarctic seaice environment. *Biogeosciences*, 9, 1137-1157.
25. Heukelem, L. V. and Thomas, C. S. (2001). Computer-assisted high-performance liquidchromatography method development with applications to the isolation and analysis of phytoplankton pigments. *Journal of Chromatography A*, 910 (1), 31-49.
26. Hobbie, J.E., Daley, R. J., Jasper, S. (1977). Use of nucleopore filters for counting bacteria by fluorescence microscopy. *Applied and Environmental Microbiology*, 33,1225-1228.
27. Hutchins,D. A. and Bruland, K.W. 1998. Iron-limited diatom growth and Si, N uptake ratios in a coastal upwelling regime. *Nature*, 393, 561-564.
28. Jonas, R. B. (1989). Acute Copper and Cupric Ion Toxicity in an Estuarine Microbial Community. *Applied Environmental Microbiology*, 55(1), 43-49.
29. Jordi, A., Basterretxea, G., Tovar-Sánchez, A., Alastuey, A., and Querol, X. (2012). *Proceedings of National Academy of Science America*, 109(52), 21246-21249.

30. Küpper, H., Küpper, F. & Spiller, M. (1996). Environmental relevance of heavy metal substituted chlorophylls using the example of water plants. *Journal of Experimental Botany*, 47,259-66.
31. Lavaud, J. Rousseau, B. van Gorkum H. J. and A. L. (2002). Etienne, Influence of the diadinoxanthin pool size on photoprotection in the marine planctonic diatom *Phaeodactylum tricornutum*, *Plant Physiology*, 129,1398-406.
32. Laws E. A. (1991). Photosynthetic quotients, new production and net community production in the open ocean. *Deep Sea Research*, 38,143-167
33. Laws, E. A., Popp. B. N., Bidigare, R. R., Kennicutt, M. C., and Macko, S. A. (1995). Dependence of phytoplankton carbon isotopic composition on growth rate and  $[CO_2]_{aq}$ , Theoretical considerations and experimental results, *Geochimica Cosmochimica Acta*. 59. 1131-1138.
34. Laws, E. A, Bidigare, R. R and Popp, B. N. (1997). Effect of growth rate and  $CO_2$  concentration on carbon isotopic fractionation by the marine diatom *Phaeodactylum tricornutum*. *Limnology and Oceanography*, 42(7),1552-1560.
35. Letelier, M.E., Lepe, A.M., Faundez, M., Salazar, J. M. R., Aracena, P., Speisky, H. (2005). Possible mechanisms underlying copper-induced damage in biological membranes leading to cellular toxicity. *Chemical Biological Interaction*, 151,71-82.
36. Levy, J.L., B.M. Angel, J.L. Stauber, W.L. Poon, S.L. Simpson, S.H. Cheng and Jolley, D.F. (2008), Uptake and internalisation of copper by three marine microalgae, Comparison of copper sensitive and copper tolerant species. *Aquatic Toxicology*, 89, 82-93.
- Manimaran, K., Karthikeyan, P., Ashokkumar, S., Ashok Prabu, V., Sampathkumar, P. (2012). Effect of copper on growth and enzyme activities of

marine diatom, *Odontellamobiliensis*. Bulletin of Environmental Contamination and Toxicology, 88, 30-37.

38 Moffet, J. W. and Brand, E. L. (1996). Production of strong extracellular Cu chelators by marine cyanobacteria to Cu stress. Limnology and Oceanography, 41 (3), 388-395.

39. Morel, F. M. M., Reinfelder, J. R., Roberts, S. B., Chamberlain, C. P., Lee, J. G. and Yes, D. (1994). Zinc and carbon co-limitation of marine phytoplankton. Nature, 369, 740-742

40. Morel, N. M. L., Rueter, J. G. and Morel, F. M. M. (1978) Copper toxicity to *Skeletonema costatum*, Journal of Phycology, 11, 43-48.

41. Morelli, E., Scarano, G. (2004) Copper-induced changes of non-protein thiols and antioxidant enzymes in the marine microalga *Phaeodactylum tricornutum*, Plant Science, 167, 289-296.

42. Morillo, J., Usero, J., Gracia, I. (2004). Heavy metal distribution in marine sediments from the southwest coast of Spain. Chemosphere, 55, 431-442

43. Nassiri, Y., J. L. Mansot, J. Wery, T. Ginsburger-Vogel, and J. C. Amiard (1997).

Ultrastructural and electron energy loss spectroscopy studies of sequestration mechanisms of Cd and Cu in the marine diatom *Skeletonema costatum*. Achieves of Environmental Contamination and Toxicology, 33, 147-155.

44. Nguyen-Deroche et al. (2009). Effects of copper on growth and photosynthesis in marine diatoms, a comparison between species from two different geographical areas. Cryptogamie Algologie, 30(2), 97-109.

45. Paasche, E. (1973). Silicon and Ecology and Marine planktonic diatoms. 1. *Thalassiosira pseudonana* (*Cyclotella nana*) grown in chemostats with silicate as the limiting nutrient. Marine Biology, 19, 117-126.

46. Paul, J. T., Ramaiah, N., Gauns, M., Fernandes, V. (2007). Predominance of few diatom species among highly diverse microphytoplankton assemblages in the Bay of Bengal Marine Biology, 152(1), 63-75.
47. Peers, G., Quesnel, S.A. and Price, N.M. (2005). Copper requirements for iron acquisition and growth of coastal and oceanic diatoms. Limnology and Oceanography, 50(4), 1149-1158.
48. Pérez, P., Beiras, R., Fernandez, E. (2010). Monitoring copper toxicity in natural phytoplankton assemblages, application of fast repetition rate fluorometry. Ecotoxicology and Environmental Safety, 73, 1292-1303.
49. Rau, G.H., Riebesell, U. and Wolf-Gladrow, D. (1996). A model of photosynthetic <sup>14</sup>C fractionation by marine phytoplankton based on diffusive molecular CO<sub>2</sub> uptake. Oceanographic Literature Review, 11(43), 1094,
50. Rejomon, G., Balachandran, K. K., Nair, M. Joseph, T., Dineshkumar, P. K. Achuthankutty, C. T., Nair, K. K. C., Pillai, N. G. K. (2008). Trace metal concentrations in zooplankton from the eastern Arabian Sea and western Bay of Bengal. Environmental Forensics, 9, 22-32.
51. Rejomon, G., Dinesh Kumar, P.K., Nair M., Muraleedharan, K. R. (2010). Trace metal dynamics in zooplankton from the Bay of Bengal during summer monsoon. Environmental Toxicology, 25(6), 622-633.
52. Richoux, N. B., Froneman, P. W. (2009). Plankton trophodynamics at the subtropical convergence, Southern Oceans. Journal of Plankton Research, 31(9), 1059-1073.
53. Rijstenbil, J.W., Sandee, A., Van Drie, J., Wijnholds, J. A. (1994), Interaction of toxic trace metals and mechanisms of detoxification in the planktonic diatoms *Ditylum brightwellii* and *Thalassiosira pseudonana*, FEMS Microbiology Review, 14, 387-396.



54. Rueter, J. G., Chisholm, S. W., Morel, F. M. (1981). The effects of Cu toxicity on silicic acid uptake and growth in *Thalassiosira pseudonana* (Bacillariophyceae). *Journal of Phycology*, 17, 270-278.
55. Sabatini, S. E., Juárez, A. B., Eppis, M. R., Bianchi, L., Luquet, C. M., Rios de Molina, C. (2009). Oxidative stress and antioxidant defenses in two green microalgae exposed to copper. *Ecotoxicology and Environmental Safety*, 72(4), 1200-1206.
56. Shaik, A. U. R., Biswas, H., Reddy, N.P.C, Rao, S. V., Bharathi, M. D. Subbaiah, Ch.V.(2015). Time series monitoring of water quality and microalgal diversity in a tropical bay under intense anthropogenic interference (SW coast of Bay of Bengal, India). *Environmental Impact Assessment and Review*, 55, 169-181.
57. Sharp, J. H. (1991). Total mass and particulate carbon, nitrogen and phosphorus. In, Hurd, D. C., Spencer, D. W. (eds) *Marine Particles, analysis and Characterization*. Geophysical monograph. American Geophysical Union. Washington, D. C. 87-91.
58. Sinningh-Damste, J. S., Rampen, S., Irene, W., Rupstra, C., Abbas, B., Muyzer, G., and Schouten, S. (2003). A diatomaceous origin for long-chain diols and mid-chain hydroxy methyl alkanoates widely occurring in Quaternary marine sediments, Indicators for high-nutrient conditions. *Geochimica Cosmochimica Acta*, 67, 1339-1348.
59. Stauber, J. L and Florence T. M. (1985). The influence of iron on copper toxicity to the marine diatom *Nitzschia closterium* (Ehrenberg) W. Smith. *Aquatic Toxicology*, 6, 297-305.
60. Stauber, J. L, and Florence, T. M. (1987). The mechanism of toxicity of ionic copper and copper complexes to algae. *Marine Biology (Berlin)*, 94, 511-519.

61. Steeman-Nielsen, E. and Kamp-Nielsen L. (1970). Influence of deleterious concentrations of copper on the growth of *Chlorella pyrenoidosa*. *Physiologia Plantarum*, 23, 828-840.
62. Stoecker, D.K., Sunda, W.G. and Davis, L.H. (1986). Effects of copper and zinc on twoplanktonic ciliates. *Marine Biology*, 92(1),21-29.
63. Strickland, J. D., Parsons, T. R. (1972). A practical handbook of Seawater analysis, Bulletin of Fisheries Research Board of Canada, 167.
64. Stryer, L. (1988). *Biochemistry*, 3rd ed. Freeman.
65. Sunda, W.G. and Huntsman, S. A. (1983). Effects of competitive interaction between manganese and copper on cellular manganese and growth in estuarine and oceanic species of the diatom *Thalassiosira*. *Limnology Oceanography*, 28,924-934.
66. Sunda, W.G. and Huntsman, S.A. (1998). Interactions among Cu, Zn and Mn in controlling cellular Mn, Zn, and growth rate in the coastal alga *Chlamydomonas*. *Limnology and Oceanography*, 43 (6), 1055-1064.
67. Suzuki, R. and Ishimaru, T. (1990). An improved method for the determination of phytoplankton chlorophyll using N,N-dimethylformamide. *Journal of Oceanography*, 46,190-194.
68. Watanabe, T. and Kobayashi, M. (1988). Chlorophyll functional Moleculesin photosynthesis Molecular composition in vivo and Physical chemistry in vitro. Special Articles on Coordination Chemistry of Biologically important Substances, 4, 383-95.

